

Central Opioid Receptors Differentially Regulate the Nalmefene-Induced Suppression of Ethanol- and Saccharin-Reinforced Behaviors in Alcohol-Preferring (P) Rats

Harry L June^{*,1}, Rancia Cummings¹, William JA Eiler II¹, Katrina L Foster¹, Peter F McKay¹, Regat Seyoum¹, Marin Garcia¹, Shannan McCane¹, Collette Grey¹, Stephanie E Hawkins¹ and Dynesha Mason¹

¹Psychobiology of Addictions Program, Department of Psychology, Indiana University-Purdue University, Indianapolis, IN, USA

The exact opioid-sensitive receptors participating in EtOH-seeking behaviors remains unclear. Previous studies have reported higher densities of μ -opioid receptor binding in the nucleus accumbens (NACC) of P relative to NP rats; however, no differences were seen in δ -receptor binding. In contrast to the NACC, substantially lower levels of μ -receptor binding have been observed in the ventral tegmental area (VTA) of both P and NP rats, albeit no line differences have been observed. In the present study, opioid receptors in the NACC, VTA, and hippocampus were evaluated for their capacity to regulate both EtOH- and saccharin-motivated behaviors in the genetically selected alcohol-preferring (P) rat. To accomplish this, nalmefene, an opiate antagonist with preferential binding affinity for the μ -opioid receptor was unilaterally or bilaterally infused during concurrent availability of 1 h daily EtOH (10% v/v) and saccharin (0.025 or 0.050% w/v) solutions. Rats performed under a two-lever fixed ratio (FR) schedule in which four responses on one lever produced the EtOH solution, and four on a second lever produced the saccharin solution. The results demonstrated that when responding maintained by both EtOH and saccharin are matched at basal levels, unilateral (1–60 μ g) or bilateral (0.5–10 μ g) microinjections of nalmefene into the NACC produced selective dose-dependent reductions on responding maintained by EtOH. Unilateral (40, 60 μ g) and bilateral (10 μ g) VTA infusions were also observed to selectively reduced EtOH responding; however, greater nalmefene doses were required and the magnitude of suppression on EtOH responding was markedly less compared with the NACC. The greater sensitivity of nalmefene to suppress EtOH responding in the NACC is likely due to the greater number of opioid receptors in the NACC relative to the VTA. Only bilateral infusion of the 40 μ g dose in the NACC and VTA suppressed responding maintained by both EtOH and saccharin. In contrast, intrahippocampal infusions dose dependently suppressed EtOH- and saccharin-maintained responding over a range of doses (1–20 μ g). The present study provides evidence that nalmefene suppresses EtOH-motivated behaviors via blockade of opioid receptors within the NACC and VTA, and under various dose conditions both reinforcer and neuroanatomical specificity can be observed.

Neuropsychopharmacology (2004) 29, 285–299, advance online publication, 5 November 2003; doi:10.1038/sj.npp.1300338

Keywords: alcohol-preferring (P) rats; ethanol-motivated behaviors; alcohol reinforcement; nucleus accumbens; ventral tegmental area; mesolimbic; opioid receptors

INTRODUCTION

The endogenous opioid systems provide the brain with mechanisms that regulate not only opioid addiction, but the addiction of a variety of abused drugs. Unraveling the mechanisms of the endogenous opioid systems have provided valuable insight into many of these addictive behaviors, particularly in relation to alcohol dependence (Herz, 1997). While a variety of neuronal systems have been

shown to play a role in regulating EtOH-seeking behavior (McBride and Li, 1998; McBride *et al*, 1993) few have been linked as closely to the positive reinforcing properties of EtOH as the endogenous opioid systems (for a review, see Froehlich and Li, 1993; Herz, 1997; Reid and Hubbell, 1992). This conclusion is supported by pharmacological studies showing that selective (eg δ and μ) and nonselective (eg naloxone, naltrexone, nalmefene) opioid antagonists can decrease EtOH-seeking behaviors in a variety of preclinical animals models, suggesting that the endogenous opioid systems mediate in part, EtOH-reward processes (Herz, 1997; June *et al*, 1999; June *et al*, 1998; Krishnan-Sarin *et al*, 1995a). Similarly, it is also well established that the nonselective opioid antagonists naltrexone and nalmefene attenuate alcohol drinking behavior, control craving, and prevent relapse in alcohol-dependent humans (O'Malley

*Correspondence: HL June, Department of Psychology, LD 124, IUPUI, 402 North Blackford Street, Indianapolis, IN 46202-3275, USA, Tel: +1 317 274 6755, Fax: +1 317 274 6756, E-mail: hjune@iupui.edu
Received 24 September 2002; revised 02 September 2003; accepted 15 September 2003

Online publication: 25 September 2003 at <http://www.acnp.org/citations/Npp09250302351/default.pdf>

et al, 1992; Volpicelli *et al*, 1992; Mason *et al*, 1994, 1999) albeit, the effectiveness of these agents has been limited by problems with compliance (Volpicelli *et al*, 1997; Johnson and Ait-Daoud, 2000; Kranzler, 2000).

The exact neurobiological substrate(s) participating in EtOH's rewarding effects, however, are not well understood, in part because the majority of the preclinical studies using opioid antagonists have been conducted using systemic drug injections (Froehlich and Li, 1993; Herz, 1997; Reid and Hubbell, 1992; Schwartz-Stevens *et al*, 1992). While it has been assumed that the action of opioid antagonists on EtOH's reinforcing properties are mediated centrally (McBride and Li, 1998), this has only been documented in a few published reports in the literature (Heyser *et al*, 1999; Hyytiä and Kiianmaa, 1997).

Previously, Heyser *et al* (1999) showed that local injection of methylnaloxonium into the nucleus accumbens (NACC) and amygdala blocked EtOH self-administration in outbred Wistar rats. An earlier study by Hyytiä and Kiianmaa (1997) evaluated the capacity of the μ -selective antagonist, CTOP, and the δ_1 -receptor antagonist, naltrindole to reduce EtOH-maintained responding when infused into the NACC, basolateral amygdaloid nucleus (BLA), and ventral tegmental area (VTA) in Wistar rats. Their results showed that CTOP suppressed EtOH responding only when injected in the BLA, while naltrindole suppressed EtOH responding following infusion into both the NACC and BLA. In contrast, both agents were ineffective when infused into the VTA. These results suggest that both μ - and δ -opioid receptors may differentially regulate EtOH-maintained responding in the NACC and BLA, but that opioid receptors within the VTA do not appear salient in regulating responding maintained by EtOH.

While a voluminous literature support a role for nonselective and selective opioid antagonists as potential pharmacotherapies in humans, a selective role for opioid mechanisms in alcohol reward is still a matter of debate (Brown and Holtzman, 1979; Herz, 1997; Reid and Hubbell, 1992; Schwartz-Stevens *et al*, 1992; Weiss and Koob, 1991; Weiss *et al*, 1990). Some researchers have argued that while both EtOH and other palatable reinforcers are affected by opioid antagonists, the relative magnitude of suppression is far less for water, sucrose, and saccharin solutions (see June *et al*, 1998, 1999; Heyser *et al*, 1999). Still others contend that because the endogenous opioid systems mediate natural reinforcers (ie hunger, thirst, sex, etc) it is not realistic to expect a complete separation between EtOH intake and intake of other palatable reinforcers (Herz, 1997).

Unfortunately, little if any research has systematically investigated the effects of opioid antagonists on the concurrent availability of EtOH and an alternative reinforcer in rodents, and no studies could be found that have examined the action of opioid antagonists following direct microinjection into putative EtOH reinforcing areas in such paradigms. The importance of alternative and concurrently available reinforcers in EtOH-maintained responding paradigms has been previously (Carroll *et al*, 1991; Heyman and Oldfather, 1992; Petry and Heyman, 1995; Rodefer *et al*, 1999; Samson and Grant, 1985) and recently (June, 2002) addressed. In addition, no studies could be found in the literature that have used neuroanatomical control groups to

evaluate site-specific effects on EtOH-motivated behaviors, or motivated behaviors initiated by a palatable alternative solution. Thus, the degree to which selective and non-selective opioid antagonists reduce EtOH-maintained responding/drinking is not clear. Recently, June (2002) has discussed the relative importance of using neuroanatomical control groups in microinjection studies when researchers are trying to isolate brain substrates, which regulate EtOH-motivated behaviors.

The objective of the present study was to determine the role of central opioid receptors in EtOH-motivated behaviors in the alcohol-preferring (P) rat. To accomplish this, nalmefene, a high affinity ($IC_{50} \sim 1$ nM) (Michel *et al*, 1985) nonselective opioid antagonist was evaluated following unilateral and bilateral microinjection into the NACC and VTA. The unilateral infusions were performed to determine the minimal level of opioid receptor occupancy required to observe modulation of EtOH- or saccharin-motivated behaviors. The degree of neuroanatomical specificity produced by nalmefene was examined following bilateral injections into the CA1 and CA3 hippocampal areas. The hippocampal fields contain substantial levels of μ - and δ -opioid receptors (Mansour *et al*, 1987, 1995; McBride and Li, 1998); however, the opioid receptors in the hippocampus have not been reported to play a role in alcohol reinforcement. The selectivity of nalmefene on consummatory responding was evaluated by determining nalmefene's effects in P rats whose response rates for EtOH (10% v/v) and saccharin solutions (0.05% w/v) were matched at basal levels.

EXPERIMENTAL PROCEDURES

Animals

Female, selectively bred P rats ($N=56$) from the S47 and S48 generations (Lumeng *et al*, 1995) were approximately 3–4 months of age at the beginning of the experiment. No effects of estrous cycle have been observed on EtOH drinking patterns in female P rats (McKinzie *et al*, 1996), and female rats maintain their body weights within a range that allows for more accurate stereotaxic placement than male P rats (Nowak *et al*, 1998). During the initial phase of the study, the rats weighed between 204 and 346 g. Animals were individually housed in wire-mesh stainless-steel cages or plastic tubs. The vivarium was maintained at an ambient temperature of 21°C and was on a normal 12 h light cycle (lights on at 0700). All rats were provided *ad libitum* access to food and water, except during the first 2 days of the training phase wherein rats were fluid deprived 23 h daily (see below). Thereafter, rats were maintained on *ad libitum* food and water. All training and experimental sessions took place between 0900 and 1500. All procedures were conducted in strict adherence with the *NIH Guide for the Care and Use of Laboratory Animals*.

Drugs and Solutions

EtOH (10% v/v) and saccharin solutions (0.05% g/v) were prepared for the operant chamber as previously described (June *et al*, 1998, 1999). Nalmefene, [17-*N*-cyclopropylmethyl-3,14b-dihydroxy-4,5a-epoxy-6-methylene-morphinan

hydrochloride], the 6-methylene derivative of naltrexone was donated as a gift by IVAX Pharmaceuticals (Miami, FL).

Behavioral Testing Apparatus

Behavioral testing was conducted in 15 standard operant chambers (Coulbourn Instruments) equipped with two removable levers and two dipper fluid delivery systems enclosed in sound-attenuated cubicles. All dipper presentations provided a 1.5-s access to a 0.1-ml dipper, followed by a 3-s time out period. Above each lever, three stimulus lights (red, green, and yellow) were present, and a stimulus delivery/reinforcer was indicated by illumination of the middle (green) stimulus light. Responses were recorded and reinforcements were controlled by two Dell computers using the 4.0 Coulbourn L2T2 operant software package.

Behavioral Training and Testing Procedures

Rats were trained to orally self-administer EtOH in daily 60 min sessions using previously described procedures (June *et al*, 1998, 1999; June, 2002). Briefly, rats were initially placed on a 23 h fluid restriction schedule for the first 2 days of training. During this period, they were trained to respond to a 0.1% (w/v) saccharin solution on a continuous reinforcement schedule (ie fixed ratio (FR1) schedule). On day 3, following successful acquisition of responding, water was available *ad libitum* for the remainder of the experiment. On the fourth day of the training phase, EtOH (5% v/v) was added to the 0.1% (w/v) saccharin solution, and over the next 5 days, EtOH was gradually increased stepwise to 10% (v/v) (ie 5, 7, 9, 10, 10%), while saccharin decreased (0.1, 0.075, 0.05, 0.0125, 0.0%). After a period of stabilization on the concurrent FR1 schedule for EtOH and water, the response requirement was increased to a concurrent FR4 schedule. Following stabilization on the concurrent FR4 schedule, a series of preliminary studies were conducted to determine the saccharin concentration that produced response rates similar to that of EtOH (see June *et al*, 1998, 1999; June, 2002). Of the saccharin concentrations tested, the 0.025 and 0.05% w/v concentrations produced rates of responding that were similar to EtOH in the rats tested. Stabilization on the concurrent FR4 schedule for EtOH and the saccharin concentration (0.05 or 0.025% w/v) was conducted for 3 additional weeks prior to beginning the drug treatment phase. Responding was considered stable when responses were within +20% of the average responses for 5 consecutive days. The position of the levers and associated dippers for *each* reinforcer was alternated on *each* session to avoid the establishment of lever preference under all concurrent schedules.

Rationale for Selection of Brain Sites

The NACC, VTA, and hippocampus (CA1 and CA3 fields) were the brain sites selected to evaluate the actions of nalmefene on EtOH-maintained responding. The NACC was selected as the primary brain substrate of interest since the majority of the previous literature has implicated the NACC as a substrate in which several types of opioid receptors

regulate alcohol drinking behavior (Koob *et al*, 1998; McBride and Li, 1998). It was also selected as the primary substrate since moderate-high densities of different types of opioid receptors have been reported in the NACC (Mansour *et al*, 1987; McBride and Li, 1998; McBride *et al*, 1998a). The VTA was selected as the second substrate since it has been consistently reported that differences in endogenous opioids and opioid receptors in this locus might contribute to alcohol preference in some alcohol-selected rodent lines (for a review, see McBride and Li, 1998). However, opioid-receptor density within the VTA has been reported to be very low (Mansour *et al*, 1987; McBride *et al*, 1998a). Further, previous research has shown that opioid receptors in the VTA play a minor role in regulating EtOH-maintained responding (Hyytiä and Kiianmaa, 1997) or intake of a palatable sucrose solution (Ragnauth *et al*, 1997). Thus, equivocal reports appear to exist on the significance of the VTA in alcohol self-administration. The hippocampus was selected as the control locus since it is one of the few mesolimbic substrates that contains a significantly lower density of μ -opioid receptors in the P compared with the NP rats (McBride *et al*, 1998a). In addition, to our knowledge, opioid receptors within the hippocampus have not been reported to regulate alcohol-motivated behaviors. Hence, the negative association between hippocampal μ -opioid receptors and alcohol preference make the hippocampus an ideal neuroanatomical control locus for reinforcement studies.

Assignment to Surgical Groups

Despite extensive training, only a subpopulation of rats were capable of lever pressing concurrently for EtOH (10% v/v) and a saccharin reinforcer at relatively similar rates of responding. Hence, only rats that evidence response rates of approximately 70% or greater of the alternative reinforcer were selected to evaluate the drug treatments. Using this criterion, 63 rats were selected to participate in the study. Of these, 32 rats lever pressed for 10% (v/v) EtOH and 0.025% (w/v) saccharin at similar rates, while the remaining 31 lever pressed for 10% (v/v) EtOH and 0.05% (w/v) saccharin at similar rates. For the specific details on how rats are trained to lever press for two reinforcers at relatively similar rates see June (2002). Out of the 32 rats, 15 were then randomly assigned to the bilateral VTA group, and 15 to the bilateral NACC group. Of the remaining 31 rats, 10 were randomly assigned to the bilateral CA1 and CA3 hippocampal control group, while nine were randomly assigned to the unilateral VTA group and nine to the unilateral NACC group.

Implantation of Guide Cannulae

Guide cannulas were stereotaxically implanted unilaterally and bilaterally in the VTA (AP $-5.3/6.3$; ML $+1.6$; DV -9.2) and NACC (AP $+2.0/1.70$; ML $+1.8$; DV -7.2). Animals were implanted in the anterior (50%) to medial (50%) NACC and the anterior (50%) to posterior (50%) VTA since these areas have been shown to be associated with both natural and drug-related rewards (Kelley and Berridge, 2002; Rodd-Henricks *et al*, 2000; Nowak *et al*, 1998). The anterior VTA was defined as the VTA region at the level of the mammillary nuclei (Bregma -4.8 to

–5.6 mm). The posterior VTA is defined as the VTA region at the levels of the caudal linear (CLi) (Bregma –5.80 to –6.04), interpeduncular (Bregma –6.04 to –6.30), and pontine (Bregma –6.72 to –6.80) nuclei (see Ikemoto *et al*, 1997; Rodd-Henricks *et al*, 2000). Only bilateral hippocampal implants were performed in the CA1 and CA3 areas (AP –4.8; ML +3.0; DV –3.8). In experimental and control animals, the cannulas were aimed 1 mm above the intended brain locus. Stereotaxic coordinates were established according to the Paxinos and Watson atlas (1986). Rats were allowed to recover for 7 days before returning to training in the operant chamber.

Microinjection Procedures

When nalmefene was microinjected into a brain site it was dissolved in artificial cerebrospinal fluid (aCSF) (composition in mM: NaCl, 120; KCl, 4.8; KH₂PO₄, 1.2; MgSO₄, 1.2; NaHCO₃, 25; CaCl₂, 2.5; D-glucose, 10). When necessary, HCl acid or NaOH was added to the solutions to adjust pH levels to $\sim 7.4 \pm 0.1$. Nalmefene or aCSF was infused bilaterally for 5 min at a rate of 0.1 μ l/min using a 28-gauge injector cannula. The injector cannula was connected by polyethylene tubing to a 10 μ l Hamilton microsyringe. The injection volume delivered to each hemisphere was 0.5 μ l, while the total injection volume for both the left and right hemispheres was 1 μ l. Thus, if a rat received a drug treatment of 20 μ g, both the left and right hemispheres received 10 μ g in a volume of 0.5 μ l. Rats receiving unilateral infusions received their drug dose in a total injection volume of 1 μ l. The injections were delivered immediately before the operant session with a Harvard infusion pump (model A), during which time animals were able to move about freely in their home cages. The injection cannula extended 1 mm beyond the tip of the guide cannulae. After completion of the 5 min infusion period, the injector was left in place for an additional 1 min to allow diffusion from the needle tip.

Rationale for Selection of Doses in the NACC and VTA

The dose range of nalmefene was selected based on previous research in our laboratory evaluating the actions of unilateral and bilateral infusions in selected brain loci (Cummings *et al*, 1999). In these studies, we attempted to determine the minimal level of opioid receptor occupancy required to observe modulation of EtOH- or saccharin-motivated behaviors in a single hemisphere. Preliminary results suggested that the minimal effective dose (MED), which selectively suppressed EtOH-maintained responding following unilateral infusion in the NACC was 10 μ g, while the MED in the VTA was 20 μ g. The MED dose that selectively suppressed EtOH-maintained responding following bilateral infusions, in the NACC was 0.25 μ g (0.125 μ g/per hemisphere), while the MED in the VTA was 7.5 μ g. As expected, occupancy of more opioid receptors with bilateral infusions was substantially more effective than unilateral infusions in suppressing EtOH responding; hence, greater doses were required to observe comparable suppression with unilateral infusions. Thus, the highest nalmefene dose employed in the unilateral studies was 60 μ g, while the highest dose in the bilateral studies was 40 μ g. To control

for carryover effects in the present study, subsequent pretreatments were not administered until both EtOH- and saccharin-maintained responding had returned to their predrug baseline levels for at least 3 days, with a minimum of 3 days between all drug treatments and a maximum of 4 days.

Histology

After the completion of the behavioral testing, animals were killed by CO₂ inhalation. Cresyl violet (0.5 μ l) was injected into the infusion site, and the brain was removed and frozen. The frozen brains were sliced on a microtome at 50 μ m sections and the sections were stained with cresyl violet acetate. Infusions sites were examined under a light microscope and indicated on drawings adapted from the rat brain atlas of Paxinos and Watson (1986). Rats with improper placements were excluded from the final data analysis.

Blood Alcohol Concentration (BAC) Measurement

To ensure animals were consuming pharmacologically relevant amounts of EtOH during operant sessions, BACs were collected in a subset of animals on days animals did not receive drug treatment. After the first 20 min of an operant session, approximately 100 μ l of whole blood was collected from the rats tail tip into a heparin-coated microsample tube. The BAC samples were collected at the 20 min time point since the majority of EtOH-maintained responding typically occurs during the first half of the operant session (June *et al*, 1998, 1999; June, 2002). In addition, an attempt was made to determine the rising phase of the BAC since it has been demonstrated that the rising, but not the falling phase of the blood alcohol curve correlated significantly with the euphoric properties of EtOH (Lewis and June, 1990). After collection, the whole blood was immediately centrifuged for 5 min at 1100 rpm. Plasma samples of 5 μ l were collected with a Gilson Microman M-25 Pipette and injected directly into a GL-5 Analyzer (Analox Instruments; Luxenburg, MA). Microanalysis consisted of measuring the oxygen consumption in the reaction between the sample of alcohol and alcohol oxidase (AOD) using a Clark-type amperometric oxygen electrode. Alcohol reagent buffer solutions (pH 7.4) and alcohol oxidase enzymes were used in all samples tested. Results were calculated in units of mg/dl and printed within 20 s of each trial. To assure accuracy of measurement, each BAC sample was calibrated twice. The mean of the two samples was used as an index of the level of BAC content for a given rat. Single point calibrations were carried out using 5 μ l of aqueous 100 mg/dl (21.7 mmol/l) standard.

Statistical Analysis

The operant-maintained responding data were analyzed by a single factor repeated measures ANOVA with drug treatment (ie dose) as the independent factor. All microinjection data were analyzed following correct histological verification under light microscope. The dependent variables were EtOH- and saccharin-maintained responding. Each dependent variable was analyzed separately. *Post hoc*

comparisons between individual drug treatments were made using the *Newman-Keuls test* in all experiments. In general, drug treatment comparisons were made against the aCSF control condition and the no injection baselines (ie BL1, BL2). The BL1 condition was the average of 5 days before the animals received any drug treatment, while the BL2 condition was the average of 5 days after animals received their final drug treatment. The first day of the BL2 condition was taken 3 days after the rats final drug treatment. The BL1 and BL2 conditions were included in the design to evaluate the reliability of our operant training procedures across the entire duration of the study, and to serve as an additional control condition to compare our aCSF control treatment (see June *et al*, 2001; June, 2002). None of the baseline conditions were statistically significant from each other ($p > 0.05$). As a result, the BL1 and BL2 conditions were pooled into a single 'BL condition' in the bilateral NACC and VTA groups (Figures 1c and d). The BL conditions were not included in the unilateral studies due to the rather brief time course of the studies. By assessing the BL data, it was possible to determine any baseline shifting over the longer time periods within the bilateral studies. Correlated *t-test* was used to determine if the EtOH- and saccharin-maintained responding data were equivalent within the NACC, VTA, and hippocampal groups. For all cumulative time course analyses, data were evaluated using a two-way ANOVA with repeated measures over both time interval and dose.

RESULTS

Control Rates of EtOH- and Saccharin-Maintained Responding and BACs for Unilateral Rats

Under control conditions EtOH- and saccharin-maintained responding were similar in the NACC ($p > 0.05$) and VTA ($p > 0.05$) groups. Responding for the EtOH and saccharin reinforcers in the NACC group ($N = 5$) following the aCSF infusion was 148 ± 34 and 188 ± 39 , respectively (see Figure 1a). Responding for the EtOH and saccharin reinforcers in the VTA group ($N = 5$) following the aCSF infusion was 146 ± 35 and 192 ± 45 , respectively (see Figure 1b). For the two groups of rats, responding maintained by EtOH on the day of blood EtOH analysis ranged from 108 to 289 responses/30 min, while BACs ranged from 24 to 86 mg/dl. Intake of absolute EtOH ranged from 0.67 to 2.49 g/kg. In the NACC rats ($n = 5$), a significant correlation emerged between EtOH-maintained responding ($r = 0.68$, $p < 0.05$) and intake ($r = 0.78$, $p < 0.05$). The BACs in the VTA rats ($n = 5$) were also correlated with EtOH-maintained responding and intake ($r = 0.72$, $p < 0.05$, $r = 0.82$, $p < 0.05$, respectively).

Control Rates of EtOH- and Saccharin-Maintained Responding and BACs for Bilateral Rats

Under control conditions EtOH- and saccharin-maintained responding were similar in the bilateral NACC ($p > 0.05$) and VTA ($p > 0.05$) groups. Following the aCSF infusions, rats in the bilateral NACC group ($N = 9$) averaged 161 ± 31 responses for EtOH and 123 ± 19 responses for saccharin (see Figure 1c). Responding under baseline

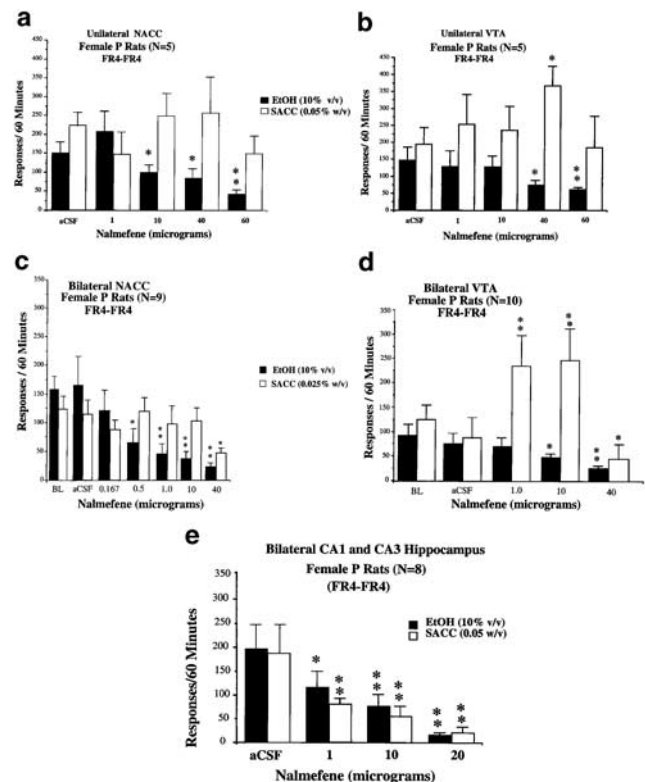


Figure 1 Effects of nalmefene (1–60 μ g) following unilateral or bilateral microinjection in the NACC (core or shell) (a, c), VTA (anterior or posterior) (b, d), or CA1 and CA3 hippocampal fields (e) on responding maintained by EtOH (10% v/v) (black bar) and saccharin (0.025 or 0.05% w/v) (white bar) during a 1 h operant session. Data are shown as mean (\pm SEM), * $P < 0.05$, ** $P < 0.01$, nalmefene vs aCSF/baseline (BL) conditions ($N = 5$ for unilateral NACC and VTA groups; $N = 9$ and 10 for bilateral NACC and VTA groups, respectively; $N = 8$ for bilateral CA1 and CA3 hippocampal groups).

conditions were similar. The bilateral NACC group consumed 5.2 ± 1.1 ml of pure EtOH and their BACs ranged from 21.6 to 94.3 mg/dl. Following the aCSF infusions, rats in the bilateral VTA group ($N = 10$) averaged 93.71 ± 21 responses for EtOH and 85 ± 27 responses for saccharin (see Figure 1d). Responding under the baseline condition was not statistically different from the aCSF condition ($p > 0.05$). The bilateral VTA group consumed 3.91 ± 1.2 ml of pure EtOH and their BACs ranged from 29.7 to 104.22 mg/dl. Rats in the bilateral hippocampal group ($N = 8$) averaged 192 ± 38 response for EtOH and 183 ± 45 responses for saccharin (see Figure 1e). The bilateral hippocampal group consumed 5.4 ± 1.4 ml of pure EtOH and their BACs ranged from 20.6 to 94.22 mg/dl. Intake of absolute EtOH ranged from 0.86 to 3.69 g/kg. In the NACC rats ($n = 7$) BACs correlated with EtOH-maintained responding and intake ($r = 0.78$, $p < 0.05$ and $r = 0.71$, $p < 0.05$, respectively). The BACs in the VTA rats ($n = 6$) were also highly correlated with EtOH-maintained responding and intake ($r = 0.81$, $p < 0.01$, and $r = 0.77$, $p < 0.01$, respectively). In addition, the BACs (ie mg/dl) in the hippocampus rats ($n = 6$) correlated significantly with EtOH-maintained responding and intake ($r = 0.79$, $p < 0.01$ and $r = 0.72$, $p < 0.01$, respectively).

Experiment 1 (NACC) Unilateral Study

Histology. Figure 4a illustrates the histologies for the five animals with correct unilateral placements in the NACC (solid circles). Of the correctly placed rats, three had their cannulae in the anterior NACC (Bregma: 2.70, 2.20), while the two remaining rats had more medial to posterior placements (Bregma: 1.70, 1.20). Representative unilateral photomicrographs (Figures 5d and e) are depicted with the bilateral photomicrographs.

Microinfusions. Figure 1a (black bar) shows rates of responding maintained by EtOH following unilateral microinjection of the 1–60 μ g doses of nalmefene into the NACC core or shell. Compared with the aCSF control condition responding was reduced with the 10–60 μ g doses by 44–83% of control levels resulting in a highly significant main effect of drug treatment ($F(4, 16) = 12.13$, $p < 0.0001$). In contrast, unilateral nalmefene produced little effects on saccharin-maintained responding ($F(4, 16) = 0.576$, $p > 0.05$) (Figure 1a, white bar). The Newman-Keuls *post hoc* test confirmed that the 10–60 μ g doses significantly reduced EtOH-maintained responding ($p < 0.05$); however, no effects were seen on saccharin-maintained responding ($p > 0.05$).

Cumulative time course. As noted above, all cumulative time course data were evaluated using a two-way ANOVA with repeated measures over both time interval and dose. The cumulative 6×5 analyses were as follows for EtOH: interval ($F(5, 24) = 32.03$, $p < 0.0001$); dose ($F(4, 96) = 77.34$, $p < 0.0001$); interaction ($F(20, 96) = 18.46$, $p < 0.0001$). The cumulative 6×5 analyses for saccharin were: interval ($F(5, 24) = 16.08$, $p < 0.0001$); dose, ($F(4, 96) = 34.22$, $p < 0.0001$); interaction ($F(20, 96) = 20.45$, $p < 0.0001$). *Post hoc* analyses using the Newman-Keuls test were conducted against the aCSF control condition. Figure 2a shows that nalmefene failed to suppress EtOH-maintained responding at the 10 min interval with any tested dose ($p > 0.05$). However, at the 20–50 min intervals nalmefene suppressed EtOH-maintained responding with the 10–60 μ g doses ($p < 0.05$), albeit suppression with the 10 μ g dose dissipated by the 60 min interval ($p > 0.05$). The 40 and 60 μ g doses continued to suppress responding at the 60 min interval ($p < 0.05$). Similar to alcohol-maintained responding, Figure 2b shows that saccharin-maintained responding was not significantly altered at the initial 10 min interval relative to the control condition ($p > 0.05$). In contrast, to alcohol responding, however, the 40 μ g dose significantly elevated saccharin responding from the 20 to 50 min interval ($p < 0.05$). The 10 μ g dose did not significantly alter saccharin-maintained responding at any interval ($p > 0.05$) (Figure 2b). At the 30–60 min intervals the 1 and 60 μ g doses suppressed responding ($p < 0.05$), but neither significantly altered responding during the 20 min interval ($p > 0.05$).

Experiment 2 (Unilateral VTA Study)

Histology. Figure 4c shows the histological placements for the five animals with correct unilateral placements in the VTA. Of the correctly placed rats, three had their cannulae in the anterior VTA at the level of the mammillary nuclei

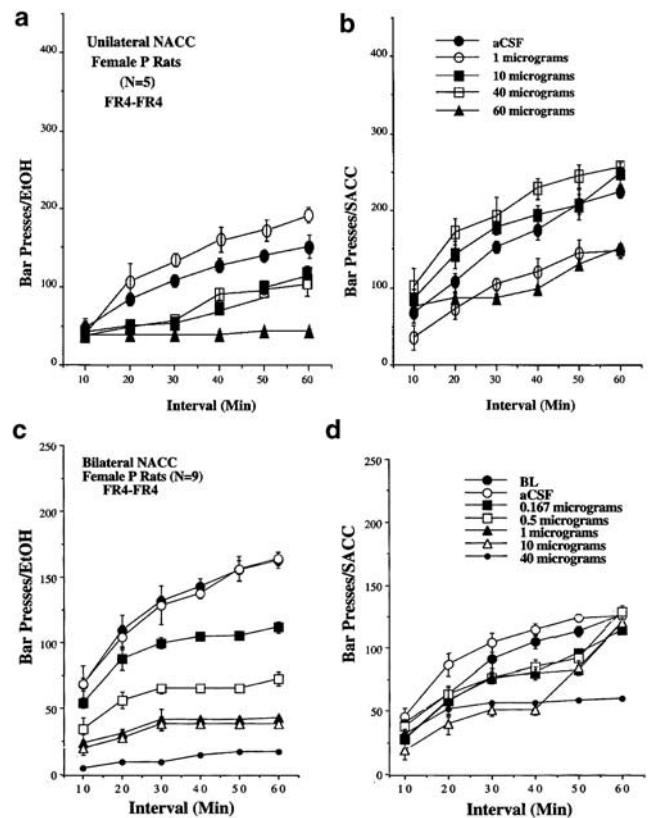


Figure 2 Cumulative time course profiles following unilateral or bilateral microinjection of nalmefene (1–60 μ g) in the NACC across the 60 min interval for EtOH (a, c) and saccharin-maintained responding (b, d) relative to the aCSF condition. The data are redrawn from Figure 1a and c. Nalmefene was without effects on EtOH responding during the initial 10 min interval following the unilateral infusions; however, the 10–60 μ g doses suppressed EtOH responding by the 20 min interval ($P < 0.05$) (a), see Results section. In contrast to the unilateral infusions, bilaterally microinjected nalmefene into the NACC produced a clear dose-dependent (eg 0.167–40 μ g) suppression on responding maintained by EtOH during the initial 10 min interval ($P < 0.05$) (c). Saccharin-maintained responding was not significantly altered at the initial 10 min interval relative to the control condition with either unilateral (b) or bilateral (d) injections ($p > 0.05$).

(Bregma: -5.20 , -5.30 , -5.60), while the two remaining rats had more posterior placements at the levels of the interpeduncular (IP) and CLi nuclei (Bregma -6.04 , -6.30). A representative unilateral photomicrograph is depicted (Figure 6c) with the bilateral histologies.

Microinfusions. Figure 1b (black bar) shows rates of responding maintained by EtOH following unilateral microinjection of the 1–60 μ g doses of nalmefene into the anterior or posterior VTA. Compared with the aCSF control condition, responding was reduced only with the 40 and 60 μ g doses. Maximal suppression was approximately 66% of control levels with the 60 μ g dose. Both doses, however, resulted in a significant main effect of drug treatment ($F(4, 16) = 3.03$, $p < 0.05$). Unilateral nalmefene produced elevations in saccharin-maintained responding with the 40 μ g dose (Figure 1b, white bar), however, the overall ANOVA only approached significance ($F(4, 16) = 2.34$, $p > 0.10$). *Post hoc* test confirmed that the 40 and 60 μ g doses significantly reduced EtOH-maintained responding

($p < 0.05$), while the 40 μg dose elevated responding maintained by saccharin ($p < 0.05$).

Cumulative response profile. The cumulative 6×5 analyses were as follows for EtOH: interval ($F(5, 24) = 19.45$, $p < 0.0001$); dose ($F(4, 96) = 8.56$, $p < 0.0001$); interaction ($F(20, 96) = 12.79$, $p < 0.0001$). Nalmefene failed to alter alcohol-maintained responding at the initial 10 min interval ($p > 0.05$) (Figure 3a). However, nalmefene suppressed EtOH-maintained responding at the 30 min interval with the 10 and 60 μg doses ($p < 0.05$). With the 40 μg dose the

suppression was not apparent until the 40 min interval ($p < 0.05$). At the 50–60 min intervals, the 10, 40, and 60 μg doses continued to suppress EtOH responding ($p < 0.05$). The cumulative 6×5 analyses for saccharin were: interval ($F(5, 24) = 23.56$, $p < 0.0001$); dose ($F(4, 96) = 56.78$, $p < 0.0001$); interaction ($F(20, 96) = 16.89$, $p < 0.0001$). Similar to the EtOH data, nalmefene failed to alter saccharin-maintained responding at the initial 10 min interval ($p > 0.05$) (Figure 3b). In contrast, the 1 and 40 μg doses elevated saccharin-maintained responding from the 20 to 60 min intervals ($p < 0.05$). In addition, the 10 μg dose also elevated saccharin-maintained responding at the 40 and 50 min intervals ($p < 0.05$), while the 60 μg dose failed to alter responding at any time interval ($p > 0.05$).

Experiment 3 (Bilateral NACC Study)

Histology. Figure 4b illustrates the histological placements for the bilateral NACC rats. Of the correctly placed rats, six rats had their cannulae in the anterior NACC (Bregma: 2.70, 2.20 mm), while two rats had more medial to posterior NACC placements (Bregma: 1.70, 1.00 mm). One rat, however, had a single cannula in the anterior NACC (Bregma: 2.70), while the other was located in the posterior NACC (Bregma: 1.00 mm). Representative photomicrographs are depicted in Figures 5a–c illustrating three pairs of bilateral guide cannula tracks.

Microinfusions. Figure 1c (black bar) shows rates of responding maintained by EtOH following bilateral microinjection of the 0.167–40 μg doses of nalmefene into the NACC core or shell. Compared with the aCSF and no injection control conditions (eg BL (average of BL1 and BL2)), bilateral nalmefene (0.167–40 μg) produced a dose-dependent suppression on EtOH-maintained behavior ($\approx 89\%$ of control levels). (EtOH mean = 163 ± 39 ; saccharin mean = 143 ± 31). The nalmefene infusions yielded a significant main effect of drug treatment ($F(8, 64) = 12.99$, $p < 0.0001$). The Newman-Keuls *post hoc* test revealed that, except for the lowest dose (ie 0.167 μg), all nalmefene doses significantly suppressed EtOH-maintained responding compared with the aCSF and baseline conditions ($p \leq 0.05$). Figure 1c also shows rates of responding maintained by saccharin. Compared with the aCSF and no injection control conditions bilateral nalmefene in doses of 0.167–10 μg was without effect on saccharin-maintained behaviors; however, the 40 μg dose reduced responding by 59% of control levels. Unlike the effects on alcohol responding, the main effect of drug dose only approached statistical significance for saccharin responding ($F(8, 64) = 1.89$, $p < 0.21$). *Post hoc* test confirmed that the 1–40 μg produced a significant dose-dependent suppression on EtOH-maintained behaviors ($p < 0.05$), however, the 0.167 μg dose was not effective ($p > 0.05$) (Figure 1c). *Post hoc* test also confirmed that only the 40 μg doses significantly suppressed saccharin-maintained responding ($p < 0.05$).

Cumulative response profile. Figures 2c and d illustrate the cumulative response profiles for EtOH and saccharin, respectively, compared with the control and nalmefene treatment conditions. The cumulative 6×9 analysis was as

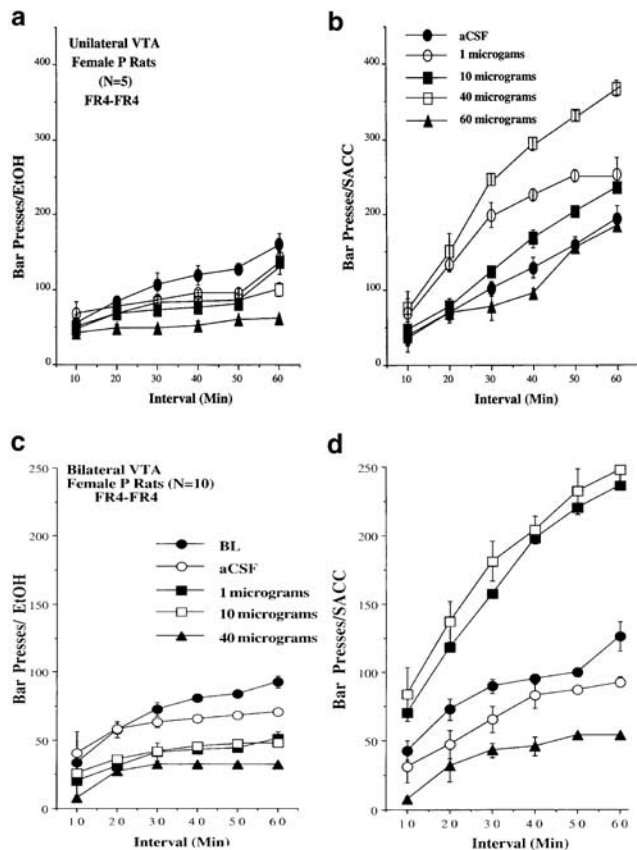


Figure 3 Cumulative time course profiles following unilateral or bilateral microinjection of nalmefene (1–60 μg) in the VTA across the 60 min interval for EtOH- (a, c) and saccharin-maintained responding (b, d) relative to the aCSF condition. The data are redrawn from Figures 1b and d. Nalmefene was without effects during the initial 10 min interval on EtOH responding following the unilateral infusions; however, all doses suppressed EtOH responding by the 30 min interval ($P < 0.05$) (a), see Results section. Unilateral nalmefene in the VTA failed to alter saccharin-maintained responding at the initial 10 min interval ($p > 0.05$) (b). However, the 1.0 and 40 μg doses elevated saccharin-maintained responding from the 20 to 60 min intervals, while the 10 μg dose also elevated saccharin-maintained responding at the 40 and 50 min intervals ($p < 0.05$) (b). Unlike the bilateral infusions in the NACC (Figure 2c), only the 40 μg dose suppressed EtOH-maintained responding during the initial 10 min interval ($p < 0.05$) (c). However, beginning at the 20 min interval, and throughout the duration of the 60 min session, the 1, 10, and 40 μg doses suppressed EtOH-maintained responding ($p < 0.05$) (c). Bilateral nalmefene in the VTA produced mild elevations with the 1 and 10 μg doses on saccharin-maintained responding at the 10 min interval ($p < 0.05$) (d), while profound elevations were seen at the 30–60 min intervals ($p < 0.01$). In contrast, the 40 μg dose suppressed saccharin-maintained responding at all intervals ($p < 0.05$), except the 20 min interval ($p > 0.05$) relative to the control conditions (d).

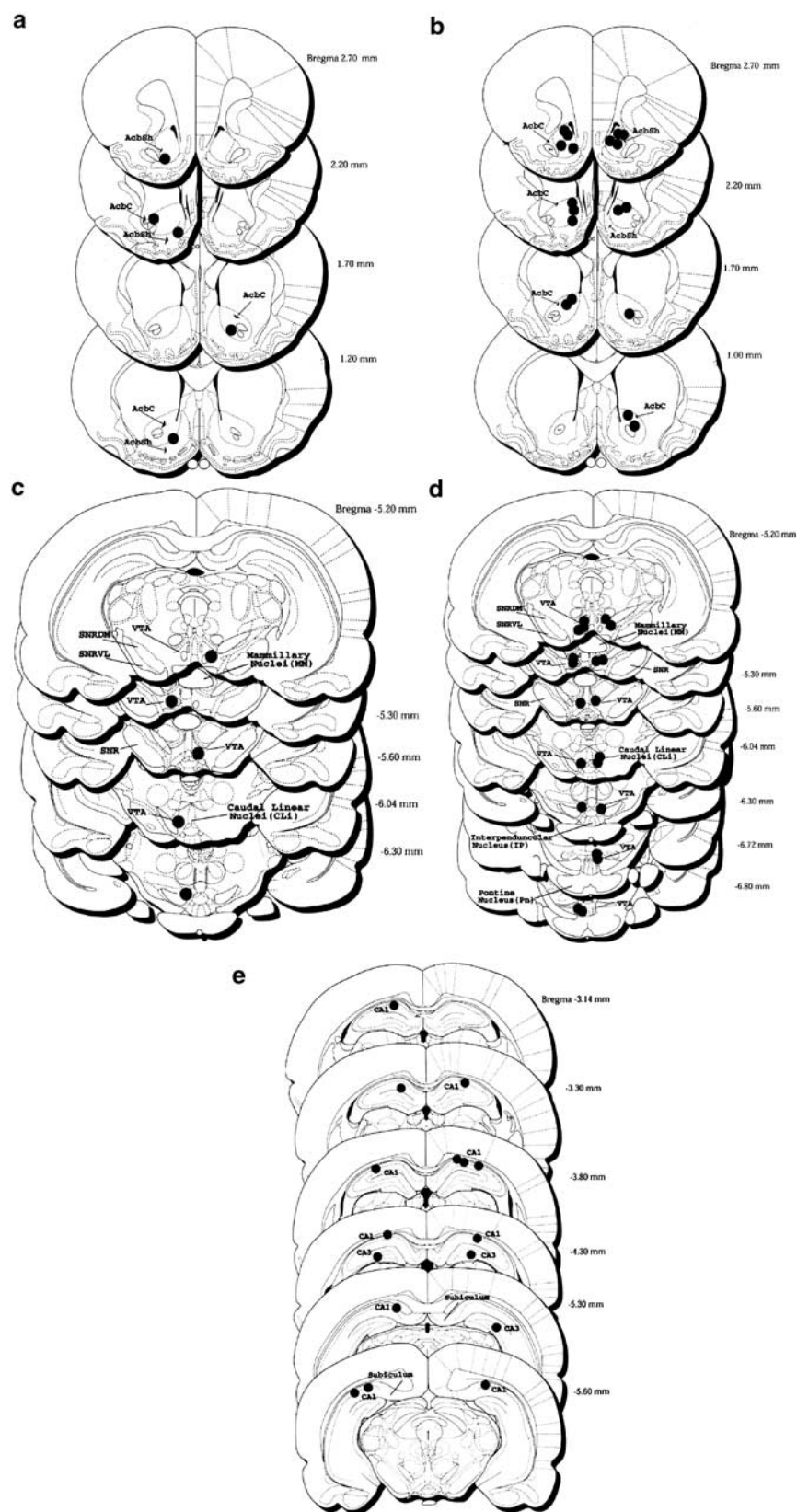


Figure 4 Reconstruction of serial coronal sections of the 'P' rat brains illustrating the unilateral ($N=5$) (a) and bilateral ($N=9$) (b) guide cannula tips in the NACC (shell and core). The unilateral ($N=5$) (c) and bilateral ($N=10$) (d) guide cannula tips in the VTA are also illustrated. Both the anterior VTA at the level of the mammillary nuclei (Bregma -4.52 to -5.60 mm) and posterior VTA at the level of the CLi/rostral interpeduncular nucleus (IPR) (Bregma -6.04) are shown. The hippocampus rats (eg neuroanatomical controls) are also depicted in the CA1 and CA3 areas (e). Rats in the unilateral groups are represented by one solid black circle in either hemisphere. However, rats in the bilateral groups are represented by one solid black circle in the left, and one in the right hemispheres. Coronal typography sections are adapted from the rat brain atlas of Paxinos and Watson (1986).

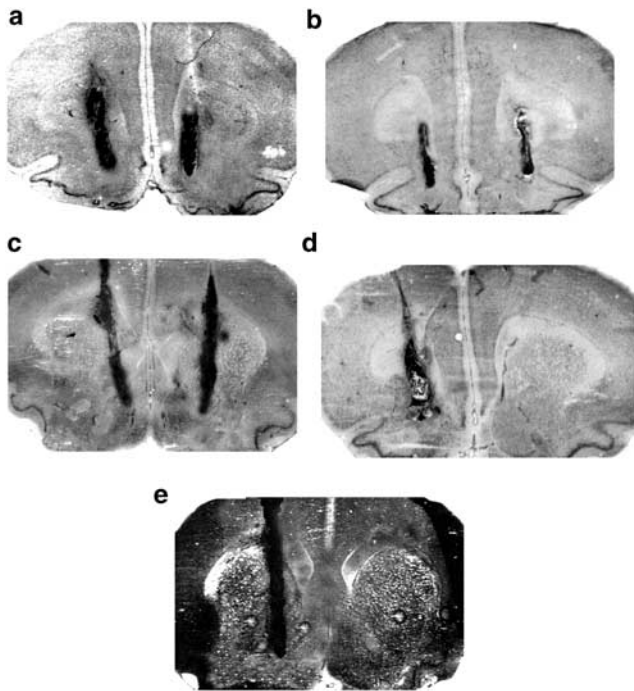


Figure 5 Representative histological photomicrographs of coronal sections for three rats with bilateral tracks (a–c) and two with a unilateral track (d, e) illustrating the anterior to medial NACC. Coronal typography was determined based on the rat brain atlas of Paxinos and Watson (1986).

follows for EtOH: interval ($F(5, 48) = 8.06$, $p < 0.0001$); dose ($F(8, 348) = 64.06$, $p < 0.0001$); interaction ($F(40, 384) = 11.76$, $p < 0.0001$). Bilateral nalmefene infusions produced a clear dose-dependent suppression (eg 0.5–40 μg) during the initial 10 min interval ($p \leq 0.05$) (see Figure 2c). This contrasted the effects observed with the unilateral infusions in the NACC and VTA groups (see Figures 2a and 3a). Furthermore, with the exception of the 0.167 μg dose at the 10 and 20 min intervals, the suppression was sustained for all doses throughout the entire 60 min session ($p \leq 0.01$). The cumulative 6×9 analyses were as follows for saccharin: interval ($F(5, 48) = 10.04$, $p < 0.0001$); dose ($F(8, 348) = 54.90$, $p < 0.0001$); interaction ($F(40, 384) = 19.45$, $p < 0.0001$). Figure 2d shows that unlike the EtOH data, none of the nalmefene infusions altered responding for saccharin at the 10 min interval ($p > 0.05$). Moreover, the 10 and 40 μg doses did not reduce saccharin-maintained responding until the 30 min interval ($p < 0.01$), with the 40 μg dose continuing to suppress responding throughout the remainder of the 60 min session ($p < 0.01$). Significant suppression was seen with the lower doses (eg 5 and 1 μg doses) only at the 40 and 50 min intervals ($p < 0.05$), albeit the suppression dissipated by the 60 min interval ($p > 0.05$).

Experiment 4 (Bilateral VTA Study)

Histology. Figure 4d illustrates the histological placements for the bilateral VTA rats. Examination of the histological placements revealed that of the correctly placed rats, five

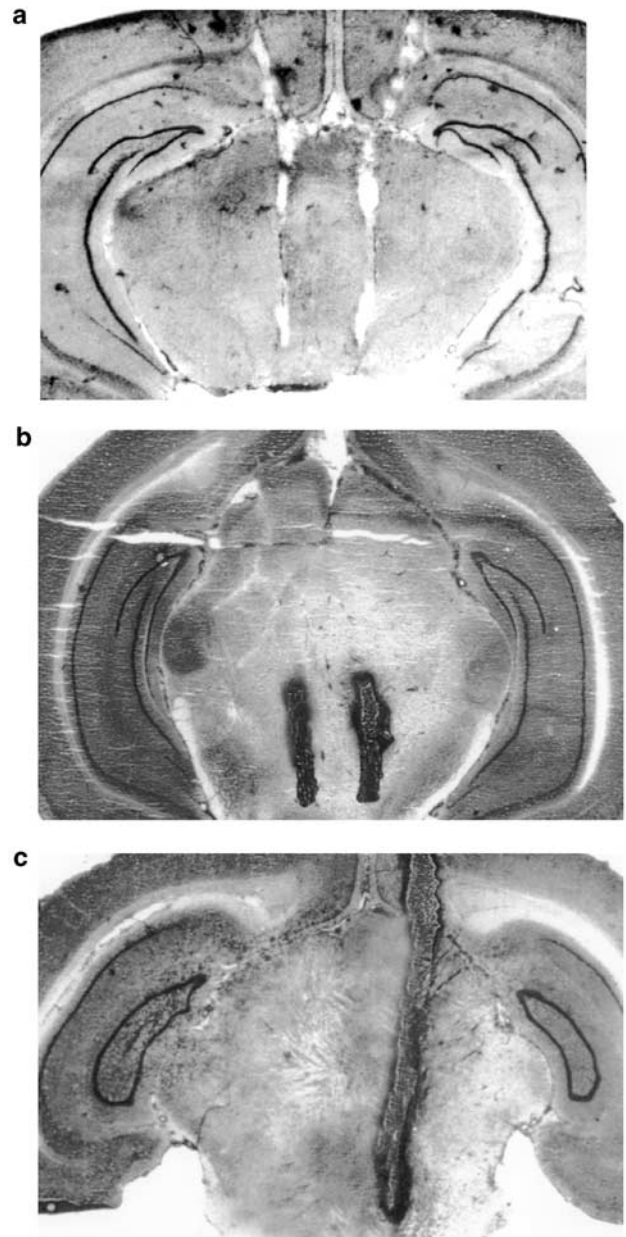


Figure 6 Representative histological photomicrographs of coronal sections for two rats with bilateral tracks in the anterior VTA (a, b), and one with a unilateral track in the posterior VTA (c). Coronal typography was determined based on the rat brain atlas of Paxinos and Watson (1986).

had both cannulae directly in the anterior VTA (Bregma -5.20 , -5.30 , -5.60), four had both cannulae in the posterior VTA (Bregma -6.04 to -6.80), while one rat had a single cannula in the anterior VTA (Bregma -5.60) and the other cannula in the posterior VTA (Bregma -6.04). Figures 6a and b depict the actual placements for '2' of the bilaterally implanted VTA rats in separate photomicrographs.

Microinfusions. Figure 1d (black bar) shows rates of responding maintained by EtOH following bilateral microinjection of the 1–40 μg doses of nalmefene into the anterior

or posterior VTA. Nalmefene produced a dose-related suppression on EtOH-maintained responding with the 1–40 μg doses. Compared with the aCSF and no injection control condition (eg BL), responding was reduced by 29–73% of control levels, resulting in a significant main effect of drug treatment ($F(4, 40) = 5.65$, $p < 0.0011$). The Newman–Keuls *post hoc* test revealed, however, that only the 10 and 40 μg doses significantly suppressed EtOH-maintained responding compared with the aCSF and baseline conditions ($p \leq 0.05$). Figure 1d (white bar) also shows rates of responding maintained by saccharin following bilateral microinjection of the 1–40 μg doses of nalmefene. Compared with the aCSF and no injection control condition (BL), responding was reduced by 66 and 34%, respectively, with the 40 μg dose. In contrast, the 1 μg dose markedly elevated saccharin-maintained responding by 204 and 177%, respectively, while the 10 μg dose elevated saccharin-maintained responding by 186 and 223%, respectively (Figure 1d). These data profiles yielded a significant main effect of drug dose ($F(4, 40) = 6.03$, $p < 0.0007$). *Post hoc* analyses confirmed that the 1 and 10 μg doses significantly elevated responding maintained by saccharin ($p < 0.01$), while the 40 μg dose significantly suppressed responding ($p < 0.05$).

Cumulative response profile. The cumulative 6×5 analyses were as follows for EtOH: interval ($F(5, 54) = 7.36$, $p < 0.0001$); dose ($F(4, 220) = 18.34$, $p < 0.0001$); interaction ($F(20, 220) = 12.56$, $p < 0.0001$). Figure 3c shows that unlike the bilateral infusions in the NACC (Figure 2c), only the 40 μg dose suppressed EtOH-maintained responding during the initial 10 min interval ($p < 0.05$). However, beginning at the 20 min interval, and throughout the duration of the 60 min session, the 1, 10, and 40 μg doses suppressed EtOH-maintained responding ($p < 0.05$). The cumulative 6×5 analyses for saccharin were: interval ($F(5, 54) = 13.35$, $p < 0.0001$); dose ($F(4, 220) = 137.66$, $p < 0.0001$); interaction ($F(20, 220) = 20.84$, $p < 0.0001$). *Post hoc* analyses confirmed that at the 10 min interval mild elevations were seen for the 1 and 10 μg doses on saccharin-maintained responding ($p < 0.05$) (Figure 3d), while profound elevations were seen at the 30–60 min intervals ($p < 0.01$). The 40 μg dose suppressed saccharin-maintained responding at all intervals ($p < 0.05$), except the 20 min interval ($p > 0.05$) relative to the control conditions.

Experiment 5 (Bilateral CA1 and CA3 Hippocampal Study)

Histology. Figure 4e shows the location of the bilateral microinjection cannulae in the CA1 and CA3 hippocampus ($N = 8$), the control locus. Examination of the histological placements revealed that five rats had both cannulae in the dorsal CA1 hippocampus, while the remaining three rats had one cannula in the dorsal CA1 hippocampus and the other in the ventral CA3 hippocampus.

Microinfusions. Figure 1e shows rates of responding maintained by EtOH (black bar) following bilateral microinjection of the 1–20 μg doses of nalmefene into the hippocampus. Compared with the aCSF control condition,

nalmefene produced a clear dose-dependent suppression on EtOH-maintained responding yielding a significant main effect of drug treatment ($F(3, 21) = 5.06$, $p < 0.0086$). Responding was reduced by 41, 61, and 91% of control levels by the 1, 10, and 20 μg doses, respectively. *Post hoc* test confirmed that all doses significantly suppressed EtOH-maintained responding compared with the aCSF condition ($p \leq 0.05$). Figure 1e also shows rates of responding maintained by saccharin. Compared with the aCSF control condition nalmefene also dose-dependently reduced saccharin-maintained responding yielding a significant main effect of drug treatment ($F(3, 21) = 3.04$, $p < 0.051$). Responding was reduced with the 1, 10, and 20 μg infusions by 56, 67, and 80% of control levels, respectively. *Post hoc* test confirmed that the 1–20 μg doses significantly suppressed saccharin-maintained responding relative to the aCSF condition ($p \leq 0.05$).

DISCUSSION

In the present study, selectively bred alcohol-preferring rats were trained to lever-press for EtOH concurrently with the availability of a palatable saccharin reinforcer. Prior to administering any drug treatment, efforts were made to match rates of responding for the two reinforcers (ie equate reward efficacy). While similar response rates for two qualitatively different reinforcers is not a guarantee of equal efficacy, when rates approximate each other at basal levels, the confound of rate may be mitigated as a factor contributing to the effect of an antagonist on EtOH-maintained responding (Samson *et al*, 1989; Carroll *et al*, 1991; Petry and Heyman, 1995; Rodefer *et al*, 1999; June, 2002).

Intra-Accumbens and Ventral Tegmental Infusions of Nalmefene Selectively Reduce Motivated Responding for EtOH when Basal EtOH and Saccharin Response Rates are Matched

A major finding of the present study was that when responding maintained by both EtOH and saccharin are matched at basal levels, microinjection of nalmefene into the NACC produced consistent dose-related and selective reductions on EtOH-maintained responding. The effects on EtOH-maintained responding were seen following unilateral as well as bilateral infusions, suggesting that only a minimal level of opioid receptor occupancy is required to observe suppression on EtOH responding within the NACC. Responding maintained by saccharin was reduced only with the highest tested dose (eg 40 μg), and when the infusions were given bilaterally. The second major finding of the present study was that selected nalmefene doses in the VTA also produced selective reductions on responding maintained by EtOH. As with the NACC infusions, responding maintained by saccharin was reduced only with the highest tested dose (eg 40 μg) and with bilateral infusions. Hence, it is likely that the suppression on saccharin responding under these conditions in both the NACC and VTA groups was due to a general suppression on consummatory behaviors.

Differential Dose and Reinforcer 'Sensitivity' Exist in the Capacity of Intra-Accumbens and Ventral Tegmental Infusions of Nalmefene to Selectively Reduce EtOH-Motivated Behaviors

The data of the present study also provide strong support for the hypothesis that a differential sensitivity exists in the capacity of centrally administered nalmefene to reduce EtOH compared with saccharin-maintained responding in both the NACC and VTA, and that a clear separation in relation to specificity of suppression on EtOH responding is possible over a broad range of doses in operant studies. We previously reported a similar separation for nalmefene following parenteral administration (June *et al*, 1998). Similar to the present study, both the saccharin and EtOH were equated at basal levels in our previous work. In the present study, the intra-accumbens and VTA modulation by nalmefene was site specific; no selectivity in suppressing EtOH-maintained responding was observed in the CA1 and CA3 hippocampal fields, even with the lowest tested dose (eg 1 μ g). Thus, opioid receptors within some (eg NACC and VTA), but not all mesolimbic substrates can selectively modulate EtOH-reinforced behaviors. Previous reports in the literature have suggested that a selectivity in reducing EtOH-motivated behaviors in rodents was difficult, equivocal, or not clearly observable (Brown and Holtzman, 1979; Herz, 1997; Schwartz-Stevens *et al*, 1992; Weiss and Koob, 1991; Weiss *et al*, 1990). However, we hypothesized that the operant contingencies under which one evaluates opiate receptor antagonists following parenteral or direct administrations into the CNS is a very important factor in observing selective antagonism on EtOH-motivated behaviors.

Nalmefene also produced a differential dose sensitivity in suppressing EtOH-motivated responding in the NACC compared with the VTA. The MED, which suppressed EtOH-maintained responding following unilateral infusion in the NACC was 10 μ g, while the MED in the VTA was 40 μ g. The MED dose following bilateral infusions, in the NACC was 0.5 μ g (0.25 μ g/per hemisphere), while the MED in the VTA was 10 μ g. EtOH responding was reduced to 56 and 85% of control levels with doses as low as 0.5 and 1 μ g, respectively. Thus, nalmefene produced a 20-fold greater potency in the NACC compared with the VTA in suppressing EtOH-maintained responding following bilateral infusions. The greater sensitivity of nalmefene in the NACC is likely due to the greater number of μ -, and possibly δ -opioid receptors in the NACC compared with the VTA (Mansour *et al*, 1987, 1995; McBride *et al*, 1998a, b; Temple and Zukin, 1987).

Nalmefene's potency also appeared to diminish substantially given unilaterally in both brain loci. These findings suggest that while unilateral injections of nalmefene with moderate-to-high doses is sufficient to produce suppression of EtOH-maintained responding, maximal receptor occupancy is achieved with bilateral infusions. Consistent with the 'low density of opioid receptor hypothesis in the VTA', Ragnauth *et al* (1997) reported that even with doses of 20–50 μ g of naltrexone the suppression on food intake following intra-VTA infusions was very small. In addition, Hyytiä and Kiianmaa (1997) reported that microinjections of

the μ -selective antagonist, CTOP, and the δ_1 -receptor antagonist, naltrindole failed to significantly alter EtOH-maintained responding when infused into the VTA. Both of the above findings appear consistent in part, with the present study. However, the differential effects between the Hyytiä and Kiianmaa (1997) and the present study could likely be due to the rat line used, dose of the opiate antagonist given, or contingencies of the operant session. Specifically, the highest bilaterally injected dose in their study was 0.5 μ g and the infusions were performed in Wistar rats, whose basal response rate was substantially lower than the P rats. The highest bilaterally injected dose in the present study with the P rats was 40 μ g. In addition, only EtOH was presented to the Wistar rats, while we equated both EtOH and a saccharin reinforcer in the current study.

Opioid Receptors in the NACC Play a More Salient Role than VTA Receptors in Modulating the Onset of EtOH-Motivated Responding

Previously, Heyser *et al* (1999) reported that local injections of methylnaloxonium into the NACC and amygdala blocked EtOH self-administration in Wistar rats during a 30 min operant session. When cumulative time course analyses were conducted, they reported that the suppression was not on the early phase (ie 0–10 min) of the operant session, but rather occurred during the middle (10–15 min) of the 30 min session. One interpretation of these data is that opioid antagonist reduced EtOH-motivated behaviors via suppressing the maintenance, but not the onset of EtOH-reward related behaviors. In the present study, we conducted a detailed cumulative time course analyses across the 60 min operant session following both unilateral and bilateral infusions. In general, our results can be summarized as follows: (1) bilateral infusion of both low and high nalmefene doses (eg 0.5–40 μ g) in the NACC reliably suppresses EtOH-motivated behaviors during the 'onset' (ie initial 10 min interval) of the operant session; (2) unilateral infusion of nalmefene doses in all brain loci fails to suppress the 'onset' of EtOH-motivated behaviors during the initial 10 min interval, however, by the end of the 20 min interval the suppression is reliably observed following both unilateral and bilateral infusions in the NACC and VTA; (3) bilateral infusion of the highest nalmefene doses (eg 40 μ g) in the VTA, but not the NACC reliably suppresses the 'onset' of saccharin-motivated behaviors; however, unilateral infusions do not alter the onset of saccharin responding. Thus, taken together, these data suggest the onset of alcohol drinking seem to be under the control of a number of variables (dose, brain loci, contingencies of the operant session, and the type of infusion (ie unilateral vs bilateral)). Nevertheless, under the conditions of the present study, our data strongly suggest that opioid receptors within the NACC appear to play a much greater role than those in the VTA in the onset of EtOH responding. In addition, opioid receptors within the NACC also appear to play a greater role in the onset of EtOH compared with saccharin responding.

Positive/Functional Association Between μ -Opioid Receptors in the NACC, but not VTA and Hippocampus and Alcohol Preference; Relation to Previous Research

Employing quantitative autoradiographic methods, McBride *et al* (1998a) reported higher densities of μ -opioid receptors in the the NACC shell and core in P relative to NP rats, however, no differences were seen in the lines between δ -receptor binding within the NACC (McBride *et al*, 1998b). In contrast to the NACC, substantially lower levels of μ -receptor binding have been observed in the VTA of both P and NP rats, albeit no line differences have been observed (McBride *et al*, 1998a). A similar differentiation of μ -opioid receptors in the NACC and VTA have also been observed in outbred rats (Mansour *et al*, 1987, 1995). It should be noted that the differences observed in the P and NP rats in the McBride *et al* (1998a) study are consistent with the higher densities of μ -opioid receptors observed in the NACC of the AA compared with the ANA rat (DeWaele *et al*, 1994). It is possible that these innate differences in binding to μ -opioid receptors may contribute to the disparate alcohol drinking in the P and AA rats. It has been suggested that the higher densities could reflect a greater number of μ -opioid receptors, which could result in 'a greater responsiveness and sensitivity of the μ -opioid receptor system to alcohol' (McBride *et al*, 1998a).

The failure of nalmefene to differentially alter EtOH- and saccharin-maintained responding in the hippocampus is noteworthy, particularly in light of the fact that compared with NP rats, P rats have a lower, not higher distribution of μ -receptor binding within the hippocampal fields (McBride *et al*, 1998a). Thus, a low, not high alcohol preference appears to be associated with enhanced μ -opioid receptor binding in the hippocampus. Recent work from our laboratory, however, has revealed that the α_5 containing GABA_A receptors in the CA1 and CA3 hippocampal fields selectively regulate EtOH, but, not saccharin-maintained responding (June *et al*, 2001). To our knowledge, only GABAergic mechanisms within the hippocampus have been shown to play such a specific role in modulating alcohol reinforcement. Nevertheless, given the failure of opioid receptors to selectively regulate EtOH-motivated behaviors within the hippocampus in the present study, combined with the negative association of EtOH preference with enhanced μ -opioid receptor binding (McBride *et al*, 1998a), we conclude that opioid receptors within the CA1 and CA3 hippocampal fields do not play a salient/specific role in regulating EtOH-motivated behaviors.

That nalmefene binds to all three major opioid receptor types (μ , δ , and κ) undoubtedly contributes to its potency in decreasing EtOH-maintained responding (Hubbell and Reid, 1990). However, nalmefene has been shown to preferentially bind to μ , κ , and δ receptors, respectively (Michel *et al*, 1985). Thus, nalmefene's potency in reducing alcohol self-administration is likely due to occupancy/activation of central μ -, rather than κ - and δ -opioid receptor subtypes. In addition, it is important to note that *in vivo* studies have shown low doses of nonselective opioid antagonists (eg naloxone, naltrexone) have been suggested to bind primarily to μ receptors (Chang and Cuatrecasas, 1981; Paterson *et al*, 1984) while higher doses (> 1.0 mg/kg) result in the occupation of δ and κ receptors (Chang and

Cuatrecasas, 1981; Leander, 1983; Leander *et al*, 1982). Thus, while nalmefene is a 'nonselective' opioid antagonist its selectivity ratio for the μ receptor markedly increases when lower doses are employed. Hence, the attenuation of EtOH self-administration in the present study particularly at the lower end of the dose response (0.5–10 μ g) may be due, primarily to an action of nalmefene at μ -opioid receptors. Similar conclusions have been reached by other investigators employing parenteral administration of non-selective opiate antagonists with humans, primates, and rodents (June *et al*, 1998, 1999; Mason *et al*, 1994; Rodefer *et al*, 1999).

Selectivity on Ingestive Behaviors: The Role of Brain Loci and Receptor Subtype; Relation to Current Research on Selective and Nonselective Opioid Antagonists

While the results of the present study suggest that microinjection of nalmefene in the NACC and VTA may preferentially attenuate the reinforcing properties of EtOH when more than one reinforcer is presented concurrently, it is important to note that selective and nonselective opioid receptor antagonists differentially reduce various forms of caloric and noncaloric food intake following central administration (Beczowska *et al*, 1993; Bodnar, 1996; Bodnar *et al*, 1995; Gosnell *et al*, 1986; Kelley *et al*, 1996; Ragnauth *et al*, 1997). However, because the saccharin solution in the present study was devoid of calories, the degree to which nalmefene preferentially attenuates motivated responding for caloric solutions when given concurrently with EtOH is not clear. Moreover, use of the concurrent operant schedule in the present study make direct comparison across studies difficult. One hypothesis that could explain the transient increases in saccharin responding may be related to a compensatory elevation in the alternative reinforcer as a result of the reduction on responding for alcohol (see June, 2002). In fact, it has been suggested that such 'compensatory increases' are further evidence of the selectivity of a particular antagonist (see June, 2002). However, it should be noted that 'compensatory increases' in the alternative reinforcer were not observed in the NACC groups. These findings suggest that 'compensatory increases' in saccharin, or other palatable reinforcers may be more likely to occur in the VTA compared with the NACC. Moreover, the VTA may be a more viable substrate in regulating both saccharin and EtOH responding, while the NACC plays a greater role in regulating EtOH-motivated behaviors.

Bodnar and colleagues (Bodnar, 1996; Ragnauth *et al*, 1997) have evaluated the effects of nonselective and selective opioid receptor antagonists following i.c.v. and direct intracerebral injections on a variety of ingestive behaviors including palatability induced feeding (ie sucrose, saccharin). These data suggest that given i.c.v., μ antagonists are effective in reducing saccharin and sucrose intake, δ receptors appear more salient in regulating saccharin reinforcement, while κ -opioid receptors may be more important in mediating sucrose reinforcement. Others have reported that bilateral infusion of naltrexone and a μ -selective antagonist directly in the NACC reduce sucrose intake; however, δ and κ antagonists were ineffective

(Bodnar *et al*, 1995; Kelley *et al*, 1996). Thus, naltrexone appears to suppress sucrose intake primarily due to blockade of μ -opioid receptors.

Ragnauth *et al* (1997) extended this research by evaluating the effects of bilateral infusion of naltrexone, μ -, δ 1-, δ 2-, and κ 1-subtype antagonists in the VTA on sucrose-induced feeding. Their data showed that only naltrexone (20–50 μ g) and the δ 2 antagonist were effective in reducing sucrose-induced feeding (albeit the maximum reduction was only 25–39%). These studies suggest that the VTA plays a 'relatively minor role' in sucrose-induced feeding, and that naltrexone may reduce sucrose intake primarily due to blockade of a variety of opioid receptor subtypes.

Finally, evidence is accumulating suggesting that an interaction exists between opioid peptides and dopamine in EtOH self-administration (Benjamin *et al*, 1993; Devine *et al*, 1993; Gonzales and Weiss, 1998; Herz, 1997). To date, the most compelling evidence to support the hypothesis is the previous work by Gonzales and Weiss (1998). Their results suggested that suppression of EtOH intake by opioid antagonists is a result of interference with 'dopamine-dependent aspects of EtOH reinforcement' within the NACC. While these preliminary results are intriguing, additional studies are warranted wherein opiate antagonists are administered centrally.

CONCLUSION

In conclusion, the present study provides evidence that nalmefene suppresses EtOH-motivated behaviors via blockade of opioid receptors within the NACC and VTA, and under various dose conditions both reinforcer and neuroanatomical specificity can be observed. The present study also provides strong data to support the hypothesis that when saccharin is used as the alternative reinforcer and rates of responding are similar to those of EtOH, EtOH is clearly more sensitive to the antagonizing effects of nalmefene following microinfusion into the NACC and VTA. That the suppression on EtOH-maintained responding occurred reliably even with unilateral injections provide further support for the hypothesis that a minimal level of opioid receptor activation is required to suppress EtOH responding, particularly within the NACC. It has also been suggested that the effectiveness of opioid antagonists is a function of palatability or preference for EtOH; thus, naltrexone may be more effective in attenuating consumption in rats with a greater preference for EtOH (Yeomans and Wright, 1991). This hypothesis, however, is mitigated by the similar response rates of the two reinforcers at basal levels. Finally, the cumulative time course profiles reveal that only bilateral nalmefene infusions in the NACC reliably suppresses EtOH-motivated behaviors during the initial phase (ie 10 min) of the operant session. Thus, the NACC may be a more pivotal substrate in regulating the onset of EtOH-motivated responding compared with its' maintenance.

ACKNOWLEDGEMENTS

We thank IVAX Pharmaceuticals (Miami, FL) for their generous donations of nalmefene. We also extend gratitude

to Drs Li (currently director of National Institute of Alcohol Abuse and Alcoholism (NIAAA)) and Lumeng and the Alcohol Research Center (Indiana University School of Medicine) for supplying the P rats. This research was supported in part by Grants R29AA10406, R01AA11555 (HLJ) from the NIAAA. Rancia Cummings was supported in part by the Minority Scholars Program of the School of Science at IUPUI. Shannan McCane and Collette Gray were supported in part by a mentorship grant from the Faculty Development Office at IUPUI. Katrina L Foster and Marin Garcia were supported in part by grant T35M from the National Heart, Lung, and Blood Institute of NIH (Short-Term Training Program for Minority Students in Biomedical Research). Katrina Foster was also supported in part by a Minority Neuroscience Fellowship from the American Psychological Association. Pete McKay was supported in part by a UROP stipend from the School of Science at IUPUI. Finally, Stephanie Hawkins was supported by the IUPUI SEED Program sponsored by the American Chemical Society.

REFERENCES

- Beczowska IW, Koch JE, Bostock ME, Leibowitz SF, Bodnar RJ (1993). Central opioid receptor subtype antagonists differentially reduce intake of saccharin and maltose dextrin solutions in rats. *Brain Res* **621**: 261–270.
- Benjamin D, Grant ER, Pohorecky LA (1993). Naltrexone reverses ethanol-induced dopamine release in the nucleus accumbens in awake, freely moving rats. *Brain Res* **621**: 137–140.
- Bodnar RJ (1996). Opioid receptor subtype antagonists and ingestion. In: Cooper SJ, Clifton PJ (eds). *Drug Receptor Subtypes and Ingestive Behaviour*. Academic Press: London. pp 127–146.
- Bodnar RJ, Glass MJ, Ragnauth A, Cooper ML (1995). General μ and κ opioid antagonists in the nucleus accumbens alter food intake under deprivation, glucoprivic and palatable conditions. *Brain Res* **700**: 205–212.
- Brown DR, Holtzman SG (1979). Suppression of deprivation-induced food and water intake in rats and mice by naloxone. *Pharmacol Biochem Behav* **11**: 567–573.
- Carroll ME, Carmona G, May SA (1991). Modifying drug-reinforced behavior by altering the economic conditions of the drug and a non-drug reinforcer. *J Exp Anal Behav* **18**: 361–376.
- Chang KJ, Cuatrecasas P (1981). Heterogeneity and properties of opioid receptors. *Fed Proc* **40**: 2729–2734.
- Cummings R, Garcia M, Mason D, McKay PF, Foster KL, June HL (1999). Differential effects of unilateral versus bilateral micro-injections of nalmefene in the nucleus accumbens and ventral tegmental area in alcohol preferring (P) rats. *Alcoholism: Clin Exp Res* **22**: 16A [Abstract # 62].
- Devine DP, Leone P, Wise RA (1993). Mesolimbic dopamine neurotransmission is increased by administration of μ -opioid receptor antagonists. *Eur J Pharmacol* **243**: 55–64.
- DeWaele JP, Kiiannmaa K, Gianoulakis C (1994). Spontaneous and ethanol-stimulated *in vitro* release of beta-endorphin by the hypothalamus of AA and ANA rats. *Alcoholism: Clin Exp Res* **18**: 1468–1473.
- Froehlich JC, Li TK (1993). Opioid peptides. In: Galanter M (ed). *Recent Developments in Alcoholism, Vol. 11: Ten Years of Progress*. Plenum Press: New York. pp 187–205.
- Gonzales RA, Weiss F (1998). Suppression of ethanol-reinforced behavior by naltrexone is associated with attenuation of the ethanol-induced increase in dialysate dopamine levels in the nucleus accumbens. *J Neurosci* **18**: 10663–10671.

- Gosnell BA, Morley JE, Levine AS (1986). Opioid-induced feeding: localization of sensitive brain sites. *277*: 119–127.
- Herz A (1997). Endogenous opioid systems and alcohol addiction. *Psychopharmacology* **129**: 99–111.
- Heyman GM, Oldfather CM (1992). Inelastic preference for ethanol in rats: an analysis of ethanol's reinforcing effects. *Psychol Sci* **3**: 122–130.
- Heyser CJ, Roberts AJ, Schulties G, Koob GF (1999). Central administration of an opiate antagonist decreases oral ethanol self-administration in rats. *Alcoholism: Clin Exp Res* **23**: 1468–1476.
- Hubbell CL, Reid LD (1990). Opioids modulate rat's intake of alcoholic beverages. In: Reid LD (ed). *Opioids, Bulimia and Alcohol Abuse and Alcoholism*. Springer-Verlag: New York. pp 145–191.
- Hyttiä P, Kiiannmaa K (1997). Intracerebral mu and delta opioid receptor antagonist decrease operant responding for alcohol in rats. *Alcohol Alcoholism* **32**: 102 [Abstract].
- Ikemoto S, Kohl RR, McBride WJ (1997). GABA_A receptor blockade in the anterior ventral tegmental area increases extracellular levels of dopamine in the nucleus accumbens of rats. *J Neurochem* **69**: 137–143.
- Johnson BA, Ait-Daoud N (2000). Neuropharmacological treatments for alcoholism: scientific basis and clinical findings. *Psychopharmacology* **149**: 327–344.
- June HL (2002). Alcohol initiation procedures in rats: methods used in evaluating potential pharmacotherapeutic agents. In: Crawley J, Gerfen C, McKay R, Rogawski M, Sibley D, Skolnick P (eds). *Current Protocols in Neuroscience* Vol. 9 John Wiley and Sons: New York. pp 1–23.
- June HL, Grey C, Warren-Reese C, Lawrence A, Thomas A, Cummings R et al (1998). The opioid receptor antagonist nalmefene reduces responding maintained by alcohol presentation: preclinical studies in alcohol preferring (P) and outbred Wistar rats. *Alcoholism: Clin Exp Res* **22**: 2174–2185.
- June HL, Harvey SC, Foster KL, McKay PF, Cummings R, Garcia M et al (2001). GABA_A-receptors containing α_5 subunits in the CA1 and CA3 hippocampal fields regulate ethanol-motivated behaviors: an extended ethanol reward circuitry. *J Neurosci* **21**: 2166–2177.
- June HL, McCane SR, Zink RW, Portoghesi PS, Li T-K, Froehlich JC (1999). The δ -opioid receptor antagonist naltriben reduces motivated responding for ethanol. *Psychopharmacology* **147**: 81–89.
- Kelley AE, Bless EP, Swanson CJ (1996). Investigation of the effects of opiate antagonist infused into the nucleus accumbens on feeding and sucrose drinking in rats. *J Pharmacol Exp Ther* **278**: 1499–1507.
- Kelley AE, Berridge KC (2002). The neuroscience of natural rewards: relevance to addictive drugs. *J Neurosci* **22**: 3312–3320.
- Koob GF, Sanna PP, Bloom FE (1998). Neuroscience of addiction [Review]. *Neuron* **21**: 467–476.
- Kranzler HR (2000). Pharmacotherapy of alcoholism: gaps in knowledge and opportunities for research. *Alcohol Alcoholism* **35**: 537–547.
- Krishnan-Sarin S, Jing S-L, Kurtz DL, Zweifel M, Portoghesi PS, Li T-K et al (1995a). The delta opioid receptor antagonist naltrindole attenuates both alcohol and saccharin intake in rats selectively bred for alcohol preference. *Psychopharmacology* **120**: 177–185.
- Leander DJ (1983). A kappa opioid effect: increased urination in the rat. *J Pharmacol Exp Ther* **224**: 89–94.
- Leander DJ, Hart JC, Lochner MA, Hynes MD, Zimmerman DM (1982). Novel phenylpiperidine opioid antagonists and partial agonists: effects on fluid consumption. *Eur J Pharmacol* **81**: 185–192.
- Lewis MJ, June HL (1990). Neurobehavioral studies of ethanol reward and activation. *Alcohol* **7**: 213–219.
- Lumeng L, Murphy JM, McBride WJ, Li T-K (1995). Genetic influences on alcohol preference in animals. In: Beglieter H, Kissin B (eds). *Genetics of Alcoholism*. Oxford University Press: New York, 165–201.
- Mansour A, Fox CA, Akil H, Watson SJ (1995). Opioid-receptor mRNA expression in the rat CNS: anatomical and functional implications. *Trends Neurosci* **18**: 22–29.
- Mansour A, Khachaturian H, Lewis ME, Akil H, Watson SJ (1987). Autoradiographic differentiation of mu, delta, and kappa opioid receptors in the rat forebrain and midbrain. *J Neurosci* **7**: 2445–2464.
- Mason BJ, Ritro EC, Morgan RO, Salvato F, Zimmer E, Goldberg G et al (1994). A double-blind, placebo-controlled pilot study to evaluate the efficacy and safety of oral nalmefene HCL for alcohol dependence. *Alcoholism: Clin Exp Res* **18**: 464.
- Mason BJ, Salvato FR, Williams LD, Rivito EC, Cutler RB (1999). A double blind, placebo-controlled study of oral nalmefene for alcohol dependence. *Arch Gen Psychiatry* **56**: 719–724.
- McBride WJ, Chernet E, McKinzie DL, Lumeng L, Li T-K (1998a). Quantitative autoradiography of mu-opioid receptors in the CNS of alcohol-naïve alcohol-preferring P and non-preferring NP rats. *Alcohol* **16**: 317–323.
- McBride WJ, Learn J, Chernet E, Lumeng L, Li TK (1998b). Regional CNS densities of mu-opioid receptors in HAD and LAD rats (Abstract no. 252). *Alcoholism: Clin Exp Res* **22**: 44A.
- McBride WJ, Li T (1998). Animal models of alcoholism: neurobiology of high alcohol-drinking behavior in rodents. *Crit Rev Neurobiol* **12**: 339–369.
- McBride WJ, Murphy JM, Gatto GJ, Levy AD, Yoshimoto K, Lumeng L et al (1993). CNS mechanisms of alcohol self-administration. *Alcohol Alcoholism* **2**(Suppl): 463–467.
- McKinzie DL, Eha R, Murphy JM, McBride WJ, Lumeng L, Li T-K (1996). Effects of taste aversion training on the acquisition of alcohol drinking in adolescent P and HAD rat lines. *Alcoholism: Clin Exp Res* **20**: 682–687.
- Michel ME, Bolger G, Weissman BA (1985). Binding of a new opiate antagonist, nalmefene, to rat brain membranes. *Methods Findings Exp Clin Pharmacol* **7**: 175–177.
- Nowak KL, McBride WJ, Lumeng L (1998). Blocking GABA_A receptors in the anterior ventral tegmental area attenuates ethanol intake of the alcohol-preferring P rat. *Psychopharmacology* **139**: 108–116.
- O'Malley SS, Jaffe AJ, Chang G, Schottenfeld RS, Meyer RE, Rounsaville B (1992). Naltrexone and coping skills therapy for alcohol dependence. *Arch Gen Psychiatry* **49**: 881–887.
- Paterson SJ, Corbett AD, Gillan MG, Kosterlitz HW, McKnight AT, Robson LE (1984). Radioligands for probing opioid receptors. *J Recept Res* **4**: 143–154.
- Paxinos G, Watson C (1986). *The Rat Brain in Stereotaxic Coordinates*. Academic Press: Sydney.
- Petry NM, Heyman GM (1995). Behavioral economic analysis of concurrent ethanol/sucrose and sucrose reinforcement in the rat: effects of altering variable-ratio requirements. *J Exp Anal Behv* **64**: 331–359.
- Ragnauth A, Ruegg H, Bodnar RJ (1997). Evaluation of opioid receptor subtype antagonist effects in the ventral tegmental area upon food intake under deprivation, glucoprivic and palatable conditions. *Brain Res* **767**: 8–16.
- Reid LD, Hubbell CL (1992). Opioids modulate rats' propensities to take alcoholic beverages. In: Naranjo CA, Sellers EM (eds). *Novel Pharmacological Interventions for Alcoholism*. Springer-Verlag: New York. pp 121–134.
- Rodd-Henricks ZA, McKenzie DL, Crille RS, Murphy JM, McBride WJ (2000). Regional heterogeneity for the intracranial self-administration of ethanol within the VTA of female Wistar rats. *Psychopharmacology* **149**: 217–224.
- Rodefer JS, Campbell UC, Cosgrove KP, Carroll ME (1999). Naltrexone pretreatment decreases the reinforcing efficacy of

- ethanol and saccharin but not PCP or food under concurrent progressive-ratio schedules in rhesus monkeys. *Psychopharmacology* **141**: 436–446.
- Samson HH, Grant KA (1985). Chlordiazepoxide effects on ethanol self-administration: dependence on concurrent conditions. *J Exp Anal Behav* **43**: 353–364.
- Samson HH, Haraguchi M, Tolliver GA, Sadeghi KG (1989). Antagonism of ethanol-reinforced behavior by the benzodiazepine inverse agonists RO15-4513 and FG 7142: relationship to sucrose reinforcement. *Pharmacol Biochem Behav* **33**: 601–608.
- Schwartz-Stevens KS, Files FJ, Samson HH (1992). Effects of morphine and naloxone on ethanol-reinforced responding in nondeprived rats. *Alcoholism: Clin Exp Res* **16**: 822–832.
- Temple A, Zukin RS (1987). Neuroanatomical patterns of μ , δ , κ , opioids receptors of rat brain as determined by quantitative *in vitro* receptor autoradiography. *Proc Natl Acad Sci USA* **84**: 4308–4312.
- Volpicelli JR, Alterman AI, Hayashida M, O'Brien CP (1992). Naltrexone and the treatment of alcohol dependence. *Arch Gen Psychiatry* **49**: 876–880.
- Volpicelli JR, Rhines KC, Rhines JS, Volpicelli LA, Alterman AI, O'Brien CP (1997). Naltrexone and alcohol dependence: the role of subject compliance. *Arch Gen Psychiatry* **54**: 737–742.
- Weiss F, Koob GF (1991). The neuropharmacology of ethanol self-administration. In: Meyer RF, Koob GF, Lewis MJ, Paul S (eds). *Neuropharmacology of Ethanol*. Birkhauser: Boston. pp 125–162.
- Weiss F, Mitchiner M, Bloom FE, Koob GF (1990). Free-choice responding for ethanol *versus* water in alcohol preferring (P) and unselected Wistar rats is differentially modified by naloxone, bromocriptine, and methysergide. *Psychopharmacology* **101**: 178–186.
- Yeomans MR, Wright P (1991). Lower pleasantness of palatable foods in nalmefene-treated human volunteers. *Appetite* **16**: 249–259.